Metal Contaminants in Largemouth Bass (Micropterus salmoides, Lacépède, 1802) from Different Origins

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ABSTRACT

The aim of this work was to evaluate the presence of some metals (Cd, Cr, Cu, Fe, Hg, Mn, Pb, Zn) on largemouth bass (Micropterus salmoides) liver and muscle tissue collected in two different areas: section of Tagus River that makes border between Portugal and Spain and in three small irrigation reservoirs located in Tagus River basin. Individuals were weighed and measured. Age were determined by examining fish scales, and sex determined by gonads observation. Samples of dorsolateral muscle, tail muscle and liver were collected to evaluate metal contaminants. We concluded that metallic concentrations of largemouth bass muscle tissues were below the maximum permissible for a safety utilization of these fishes in human nutrition.

Keywords: Heavy Metals, Cadmium, Lead, Mercury, Chromium, Muscle Tissue, Tagus River, Micropterus salmoides, Largemouth Bass.

INTRODUCTION

Largemouth bass (Micropterus salmoides, Lacépède, 1802) is a freshwater fish originating from the United States of America (USA). In the end of the XIX Century it was introduced in Portugal - Azores Islands. In main land it was introduced in 1952 (Sanches and Rodrigues, 2011).

Like in the USA, largemouth bass is one of the most popular freshwater sport’s fish in Portugal and it is very important in regional cuisine. It is much appreciated due to their exquisite flavour.

All eaten largemouth bass in Portugal are collected in large dams (Tagus and Guadiana rivers basins) and small irrigation dams (Rodrigues and Sanches, 2012). However, there’s a lack of information about metal contaminants of largemouth bass collected in Tagus River.

Heavy metals are considered the most important form of pollution in the aquatic environment because of their toxicity and because metal contents in aquatic environment have been rose by increased activities in industrial, domestic, and agricultural systems (Tuzen, 2009; Kumar et al., 2011; Mousavi et al., 2012). According Mousavi et al. (2012) the first outbreak of poisoning caused by consumption of fish contaminated with heavy metals in humans was observed in Japan - Minamata Bay in 1953, during which more than 43 local residents died and more than 700 suffered permanent disabilities after consumption of contaminated fish by industrial sewage of a factory.

For decades, Tagus River received environmental pollutants from non-point and point sources that included intensive agriculture, industrial entities, municipalities, mining activities and a Spanish nuclear power plant. This nuclear power plant is in a radius around to 150 km far from the local were largemouth bass were collected.

Heavy metals can enter from contaminated aquatic systems into fish body by different routes and accumulate in organisms’ tissues (Tuzen, 2009; Mahamadi and Chapeyama,
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Some of the metals found in fish might be essential as they play important roles in biological systems of the fish as well as in the human being. The common heavy metals that are found in fish are cadmium (Cd), chromium (Cr), copper (Cu), iron (Fe), mercury (Hg), manganese (Mn), lead (Pb) and zinc (Zn). Cu, Fe, Mn and Zn are essential metals while Cd, Cr, Hg and Pb are toxic metals (Kumar et al., 2011; Bat et al., 2017). However essential metals can have toxic and adverse effects at high concentrations (Tuzen, 2009). Metal accumulation in fish tissues has been recognized to be mainly found in liver and gills. In muscle tissue such accumulation is not so severe (Jia et al., 2017). Since fish are highly consumed by human beings and may accumulate large amounts of some metals from the water and sediments, it is important to determine the concentration of heavy metals in commercial fish in order to evaluate the possible risk of fish consumption for human health (Ekpo et al., 2008; Lopes, 2009; Tabinda et al., 2010; Khoshnoud et al., 2011; Kumar et al., 2011).

Although some studies have been done to determine largemouth bass heavy metals from small irrigation dams (Belo et al., 2007), prior to this study the status of contaminants in largemouth bass collected from the section of Tagus River that makes border between Portugal and Spain was unknown. Obviously, it is very important to know if those fishes are in safety conditions to be consumed by human beings.

The aim of this work was to evaluate some metals (Cd, Cr, Cu, Fe, Hg, Mn, Pb, and Zn) present on liver and muscle tissue of largemouth bass collected in Tagus River (TR) (N=9), compared with fish collected in three small irrigation reservoirs (IR) (N=11), located in Tagus River basin.

**MATERIAL AND METHODS**

Present research reflects a concern presented to the authors by some authorities regarding the heavy metal contamination of a popular freshwater sport’s fish very important in Portuguese regional cuisine especially in the countryside. The Tagus River is the longest river of the Iberian Peninsula with several hydroelectric dams, lentic systems where local population can fish largemouth bass.

After fishing, fishes were stored at -20°C. In sampling operations, they were slowly unfrozen, weighted, measured and aged by fish scale examination and sex determined by gonads observation. Samples for dorsolateral muscle and tail muscle were collected from the right side of the fish.

Total mercury was determined in freeze-dried samples by atomic absorption spectrometry (AAS) with thermal decomposition and gold amalgamation using an Advanced Mercury Analyser (LECO model AMA-254), as described by Costley et al. (2000). The analysis is performed directly on the sample, without digestion or specific pre-treatment, avoiding mercury losses or contamination as well as matrix interferences. At least three replicate measurements were carried out for each sample (acceptable relative standard deviation among replicates: <10%). Analytical quality control was performed by using Certified Reference Material TORT 2 - Lobster Hepatopancreas. About 200 mg of dry liver sample and 1 g of freeze dried muscle were digested with a mixture of HNO₃ (10% v/v) and H₂O₂ (30% v/v) at 100°C in a block digestion system (Digiprep MS) until complete digestion (12h). The final residue was diluted to 50 mL with HNO₃ (10% v/v) and filtered. The other heavy metals were carried out using an Inductively Coupled Plasma-Optical Emission Spectrometry (ICP-OES- Activa M, Horiba Jobin Yvon), operating at 1000 W plasma power, 15 L min⁻¹ plasma gas flow, 0.02 L min⁻¹ nebulizer air flow and 1.0 bar air pressure. The analytical wavelengths (nm) were set of the following: Cd (228.802), Cr (205.571), Cu (327.395), Fe (259.940), Mn (257.611), Pb (283.305) and Zn (213.857) (ISO, 2007). All percentages of recovery for total mercury were within the range of 80–120% (N=19). For other metals, all standard solutions used were prepared by diluting 1000 mg L⁻¹ single-element standard solutions (Prolabo, Titrinorm) in HNO₃ 0.1% (v/v). Quality control samples were from a multi-elemental standard solution of 100 mg L⁻¹ containing all analysed elements from SCP science was used. HNO₃ 65% (Merck) and H₂O₂ 30% (Prolabo) were of analytical grade and high purity Milli-Q water was used for dilutions.

Table 1 shows the characterization of the largemouth bass according TR or IR provenience. The largemouth bass TR average weight was 435.14 g (±109.15), average length 278.33 mm (±23.28), average K condition factor 1.98 (±0.09) and average age 3.11 years (±0.78) and these figures were similar (p>0.05) to largemouth bass IR average weight 410.84 g (±137.71), average length 278.36 mm (±31.13), average K
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condition factor 1.86 ±0.17 and average age 3.18 years ±0.60. Also liver weight was similar in largemouth bass TR (4.74 g ±1.29) and IR (4.25 g ±1.51) (p>0.05).

Statistical analysis was carried out with Paired-Samples T Test. For each metallic contaminant was determined mean and standard deviation (±sd). Differences in mean values were accepted as being statistically significant if p≤0.05.

Using linear regression analysis (Pearson correlation) we try to found the relationship between each metallic contaminant and largemouth bass weight, length, and K condition factor.

Table 1. TR (N=9) and IR (N=11) Largemouth Bass Average Weight, Length, K Condition Factor and Age and Liver Weight.

<table>
<thead>
<tr>
<th></th>
<th>Weigh (g)</th>
<th>Length (mm)</th>
<th>K condition factor</th>
<th>Age (year)</th>
<th>Liver weight (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Largemouth bass TR</td>
<td>435.14±109.15</td>
<td>278.33±23.28</td>
<td>1.98±0.09</td>
<td>3.11±0.78</td>
<td>4.74±1.29</td>
</tr>
<tr>
<td>Largemouth bass IR</td>
<td>410.84±137.71</td>
<td>278.36±31.13</td>
<td>1.86±0.17</td>
<td>3.18±0.60</td>
<td>4.25±1.51</td>
</tr>
<tr>
<td>Total</td>
<td>421.78±123.09</td>
<td>278.35±27.17</td>
<td>1.91±0.15</td>
<td>3.15±0.67</td>
<td>4.47±1.41</td>
</tr>
</tbody>
</table>

Note: TR – largemouth bass collected in Tagus River; IR - largemouth bass collected in three small irrigation reservoirs located in Tagus River basin; p – significant values; ns - the difference is not significant for p>0.05.

Table 2. Metal Concentration (mg.kg⁻¹ ww) on Muscle of Largemouth Bass TR (N=9) and IR (N=11).

<table>
<thead>
<tr>
<th>Elements</th>
<th>TR (mg.kg⁻¹ ww)</th>
<th>IR (mg.kg⁻¹ ww)</th>
<th>Total (mg.kg⁻¹ ww)</th>
<th>Max.(mg.kg⁻¹ ww)</th>
<th>Max. tolerable (mg.kg⁻¹ ww)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cd</td>
<td>&lt;LOQ(0.05)</td>
<td>&lt;LOQ(0.05)</td>
<td>-</td>
<td>-</td>
<td>0.05 (a)</td>
</tr>
<tr>
<td>Cr</td>
<td>&lt;LOQ(0.03)</td>
<td>&lt;LOQ(0.03)</td>
<td>-</td>
<td>-</td>
<td>1 (d)</td>
</tr>
<tr>
<td>Cu</td>
<td>0.159±0.016</td>
<td>0.148±0.017</td>
<td>0.153±0.018</td>
<td>0.181 (TR)</td>
<td>50 (b, c)</td>
</tr>
<tr>
<td>Fe</td>
<td>1.450±0.278</td>
<td>1.803±0.438</td>
<td>1.644±0.408</td>
<td>2.528 (IR)</td>
<td>50 (e)</td>
</tr>
<tr>
<td>Hg</td>
<td>0.078±0.018</td>
<td>0.223±0.173</td>
<td>0.158±0.146</td>
<td>0.546 (IR)</td>
<td>0.5 (a)</td>
</tr>
<tr>
<td>Mn</td>
<td>0.038±0.036</td>
<td>0.048±0.033</td>
<td>0.043±0.034</td>
<td>0.096 (IR)</td>
<td>4 (f)</td>
</tr>
<tr>
<td>Pb</td>
<td>&lt;LOQ(0.2)</td>
<td>&lt;LOQ(0.2)</td>
<td>-</td>
<td>-</td>
<td>0.2 (a)</td>
</tr>
<tr>
<td>Zn</td>
<td>3.760±0.263</td>
<td>4.171±0.490</td>
<td>3.986±0.447</td>
<td>5.323 (IR)</td>
<td>100 (g)</td>
</tr>
</tbody>
</table>

Note: ww – wet weight; LOQ – limit of quantification; ns - the difference is not significant for p>0.05; * - p≤0.05; (a) EU, 2008; (b) Leung et al., 2014; (c) Jia et al., 2017; (d) US EPA, 1980; (e) Elkareem, 2014; (f) Tuzen, 2009; (g) Mahamadi and Chapeyama, 2013.

Table 3. Metal Concentration (mg.kg⁻¹ ww) on Liver of Largemouth Bass TR (N=9) and IR (N=11).

<table>
<thead>
<tr>
<th>Elements</th>
<th>Liver TR (mg.kg⁻¹ ww)</th>
<th>Liver IR (mg.kg⁻¹ ww)</th>
<th>p – Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cd</td>
<td>&lt;LOQ(0.05)</td>
<td>&lt;LOQ(0.05)</td>
<td>-</td>
</tr>
<tr>
<td>Cr</td>
<td>&lt;LOQ(0.03)</td>
<td>&lt;LOQ(0.03)</td>
<td>-</td>
</tr>
<tr>
<td>Cu</td>
<td>2.05±0.60</td>
<td>5.44±6.06</td>
<td>0.114</td>
</tr>
<tr>
<td>Fe</td>
<td>74.26±23.10</td>
<td>280.98±214.07</td>
<td>0.010</td>
</tr>
<tr>
<td>Hg</td>
<td>0.03±0.006</td>
<td>0.11±0.01</td>
<td>0.050</td>
</tr>
<tr>
<td>Mn</td>
<td>1.61±0.18</td>
<td>1.50±0.53</td>
<td>0.567</td>
</tr>
<tr>
<td>Pb</td>
<td>&lt;LOQ(0.2)</td>
<td>&lt;LOQ(0.2)</td>
<td>-</td>
</tr>
<tr>
<td>Zn</td>
<td>17.38±1.51</td>
<td>18.91±5.09</td>
<td>0.399</td>
</tr>
</tbody>
</table>

Note: p – Significant values; ww – wet weight.

DISCUSSION

The mean values of Cd in both TR and IR fish were lower than EU (2008) and FAO (1983) limits (0.2 mg.kg⁻¹ ww) and lower than the figures reported by Belo et al. (2007) and Leung et al. (2014) (Micropterus salmoides) and Yousaf et al. (2012) (Wallago attu).

Cr content in both largemouth bass muscle tissues TR and IR were lower than US EPA (1989) limit (1 mg.kg⁻¹ ww) and lower than the figures...
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reported by Leung et al. (2014) (Micropterus salmoides), Tabinda et al. (2010) (Labeo rohita), Al Farraj et al. (2011) (Sepia pharaonis), Mousavi et al. (2012) (Oncorhynchus mykiss) and Elkareem et al. (2014) (Clarias lazera). There’s no difference between Cu concentration in largemouth bass TR (0.159 mg.kg⁻¹ ww ±0.016) and IR (0.148 mg.kg⁻¹ ww ±0.017).

However, the non-significant Cu higher value of TR largemouth bass could be related with the type of Spanish agriculture along Tagus River and the use of Cu for the vineyards treatment. In both cases TR and IR largemouth bass Cu were much lower than Leung et al. (2014) and Jia et al. (2017) limit (50 mg.kg⁻¹ ww) and FAO (1983) limit (10 mg.kg⁻¹ ww) and lower than the results of Belo et al. (2007) (Micropterus salmoides), Fernandes et al. (2008) (Liza saliens), Khoshnoud et al. (2011) (Scomberomorus commerson and Otolithes ruber) and Yousaf et al. (2012) (Wallago attu) but higher than Tabinda et al. (2010) (Labeo rohita) and Jia et al. (2017) (Carassius auratus, Pelteobagrus fulvidraco and Squaliobarbus curriculus).

Although the Fe concentration in both TR and IR fish muscles were much lower than Elkareem et al. (2014) limit (146 mg.kg⁻¹ ww) and lower than results reported by Belo et al. (2007) (Micropterus salmoides) and Yousaf et al. (2012) (Wallago attu), there were slightly differences (p<0.05) between Fe concentration in IR (1.803 mg.kg⁻¹ ww ±0.438) fish muscles and in TR (1.450 mg.kg⁻¹ ww ±0.278). This difference could be related with the ichthys soil around the three small irrigation dams (TR).

We did not find any significant difference between Mn in TR (0.038 mg.kg⁻¹ ww ±0.036) and IR largemouth bass muscle tissues (0.048 mg.kg⁻¹ww ±0.033) and the values of Mn in both fish TR and IR were lower than the daily upper intake limit referred by Tuzen (2009) (11 mg.day⁻¹) and much lower than values reported by Belo et al. (2007) and Leung et al. (2014) for Micropterus salmoides.

Pb contents in both TR and IR fish were lower than FAO (1983) limit (1.5 mg.kg⁻¹ ww) and EU (2008) limit (0.2 mg.kg⁻¹ ww) and lower than results reported by Belo et al. (2007), (Micropterus salmoides), Fernandes et al. (2008) (Liza saliens) and Yousaf et al. (2012) (Wallago attu).

Zn concentration was greater in IR fish muscles (4.171 mg.kg⁻¹ ww ±0.490) than in TR fish muscle (3.760 mg.kg⁻¹ ww ±0.263) (p<0.05). Both TR and IR fish muscles were found to contain Zn concentration much below limits which are 100 mg.kg⁻¹ ww (Mahamadi and Chapeyama, 2013) or 150 mg.kg⁻¹ ww (FAO, 1983). Also our figures are lower than reported by Belo et al. (2007) and Mahamadi and Chapeyama (2013) (Micropterus salmoides), Fernandes et al. (2008) (Liza saliens) and Yousaf et al. (2012) (Wallago attu) but higher than Khoshnoud et al. (2011) (Micropterus salmoides).

Finally, the average values of Hg in both fish TR and IR were below EU (2008) limit (0.5 mg.kg⁻¹ ww) and FAO (1983) limit (0.14 mg.kg⁻¹ ww). However, our results were higher than results reported by Ekpo et al. (2008) and Khoshnoud et al. (2011) (Metacembeus iconnbergii, Clarias lazera, Citharinus citharus, Tilapia zillii) but lower than total Hg concentration in largemouth bass muscle tissue (ranged from 0.12 to 0.98 mg.kg⁻¹ ww) reported by Gehringer et al. (2013). There’s a difference (p≤0.05) between Hg concentration in TR fish muscles (0.078 mg.kg⁻¹ ww ±0.018) and IR (0.223 mg.kg⁻¹ ww ±0.173). We find one IR largemouth bass with Hg value slightly higher (0.546 mg.kg⁻¹ ww) than the acceptable value (0.5 mg.kg⁻¹ ww) (EU, 2008). It was a largemouth bass with 587.3 g weight, 324 mm length and 4 years old. This fish represents 9.1% of IR largemouth bass and 5.0% of all largemouth bass collected. However, 0.546 mg.kg⁻¹ ww it is a lower value than the accepted for other fishes (1 mg.kg⁻¹ ww) which includes European eel (Anguilla anguilla), pike (Esox lucius) and other carnivorous fish (EU, 2008). An apex predator like largemouth bass has the potential, due to its trophic position, to bioaccumulate high levels of Hg (Tabinda et al., 2010; Kumar et al., 2011). Measured Hg tissue levels in largemouth bass can range from very low (<0.1 mg.kg⁻¹ ww) to extremely high (>10 mg.kg⁻¹ ww) (Rumbold and Fink 2006). According ADPH (2010) of the sixty-six mercury advisories in the Alabama coastal plain (USA), forty-nine were issued for, or included, largemouth bass. In our study 100% of TR largemouth bass and 36.4% of IR largemouth bass are Hg concentration lower than 0.1 mg.kg⁻¹ ww. In our study we found a positive correlation between Hg largemouth bass muscle concentration and length (r=0.690; p≤0.01), weight (r=0.619; p≤0.01) and fish age (r=0.554; p≤0.05) indicating that largemouth bass are accumulating
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greater amount of Hg as biggest and older they are. Like Gehringer et al. (2013), in this study total Hg concentration in muscle tissue was not related to K condition factor. However considering only largemouth bass from IR we found a very high positive correlation between Hg largemouth bass muscle concentration and length (r=0.942; p≤0.01), weight (r=0.930; p≤0.01) and fish age (r=0.873; p≤0.01). Small irrigation reservoirs are closed systems with no hydrodynamic capacity to recycle the diffuse sources of contamination.

According to our results, the concern may be greater with consumption of largemouth bass caught on small irrigation reservoirs. For this reason, we estimate the equation Hg (mg.kg⁻¹ ww) = -1.241 + 0.005 x Length (mm) (r²=0.902; p≤0.01). With this equation the fisherman can estimate the Hg concentration in largemouth bass muscle tissue after measuring the fish.

The metal concentrations on liver of largemouth bass TR and IR are shown in Table 3 and were higher than metal concentration on muscle tissue for all analyzed heavy metals, with exception of Hg. This lower level of Hg on liver relative to other tissues may be attributed to the high coordination behavior of MT polypeptides with respect to either cognate or noncognate metal ions as reported by Palacios et al. (2014). In addition, liver is the principal organ responsible for the detoxification, transportation, and storage of toxic substances and it is an active site of pathological effects induced by contamination. Higher levels of Hg in muscle than in liver are characteristic of some species, in particular on non-contaminated environments (Havelkova et al., 2008). In the samples analyzed, average levels of metal in liver are below limits, except for Fe. In general, fish liver is not consumed or it is consumed in small amounts, therefore liver is not referred in legislation. In all cases it would be safe to consume a considerable amount of fish liver.

CONCLUSIONS

Only Fe, Hg and Zn showed significant differences in largemouth bass muscle tissue from TR and IR. Largemouth bass collected in small irrigation reservoirs presented higher values than largemouth bass from Tagus River despite there were no direct contamination in irrigation reservoirs. Therefore, this indicates that Tagus River is not accumulating metals in the studied section, and can recycle the inputs from contaminants that it receives.

We concluded that metallic concentrations of largemouth bass muscle tissues collected in the section of Tagus River that makes border between Portugal and Spain and collected in small irrigation reservoirs located in Tagus River basin were below the maximum permissible for a safety utilization of these fishes as human food. Nevertheless, a good program of heavy metals control, over those and other fishes should be applied with a certain frequency to prevent future eventual contamination of fishes and the natural consequences to human health.

REFERENCES


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